

Glucocorticoids, 11 β -hydroxysteroid dehydrogenase, and fetal programming

JONATHAN R. SECKL, MARK CLEASBY, and MOFFAT J. NYIRENDA

Molecular Medicine Center, University of Edinburgh, Western General Hospital, Edinburgh, Scotland, United Kingdom

Glucocorticoids, 11 β -hydroxysteroid dehydrogenase, and fetal programming. Epidemiological studies in many distinct human populations have associated low weight or thinness at birth with a substantially increased risk of cardiovascular and metabolic disorders, including hypertension and insulin resistance/type 2 diabetes, in adult life. The concept of fetal “programming” has been advanced to explain this phenomenon. Prenatal glucocorticoid therapy reduces birthweight, and steroids are known to exert long-term organizational effects during specific “windows” of development. Therefore, we hypothesized that fetal overexposure to endogenous glucocorticoids might underpin the link between early life events and later disease. In rats, birthweight is reduced following prenatal exposure to the synthetic glucocorticoid dexamethasone, which readily crosses the placenta, or to carbenoxolone, which inhibits 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2), the physiological fetoplacental “barrier” to endogenous glucocorticoids. Although the offspring regain the weight deficit by weaning, as adults they exhibit permanent hypertension, hyperglycemia, and increased hypothalamic-pituitary-adrenal axis activity. Moreover, physiological variations in placental 11 β -HSD2 activity near term correlate directly with fetal weight. In humans, 11 β -HSD2 gene mutations produce a low birthweight, and some studies show reduced placental 11 β -HSD2 activity in association with intrauterine growth retardation. Moreover, low birthweight babies have higher plasma cortisol levels throughout adult life, indicating that hypothalamic-pituitary-adrenal axis programming also occurs in humans. The molecular mechanisms of glucocorticoid programming are beginning to be unraveled and involve permanent and tissue-specific changes in the expression of key genes, notably of the glucocorticoid receptor itself. Thus, glucocorticoid programming may explain, in part, the association between fetal events and subsequent disorders in adult life.

Extensive epidemiological studies suggest that factors operating in early life are important determinants of the risk of common and interassociated cardiovascular and metabolic disorders of adult life. Data from several distinct populations in the Europe, Asia, Australia, and North America have shown that low birthweight or thinness at birth strongly predicts the subsequent occurrence

of hypertension, hyperlipidemia, insulin resistance, type 2 diabetes, and ischemic heart disease deaths in adult life [1–13]. Classic adult lifestyle risk factors (smoking, alcohol, obesity, social class) appear to be additive to these early life effects [3]. Importantly, the relationships represent birthweights within the normal range, rather than severe intrauterine growth retardation, multiple babies, or very premature babies. Indeed, the smaller of twins at birth apparently has the higher blood pressure in later life [14]. Postnatal “catch-up” growth is also a risk factor for subsequent hypertension, ischemic heart disease, and insulin resistance [3, 9, 14, 15], perhaps suggesting that smallness at birth due to environmental influences restraining intrauterine growth are of importance. The “early life” associations appear also to be important predictors of later disease. In Preston, a small baby with a large placenta had a relative risk of adult hypertension three times that of a large baby with a normal placenta [16]. In 22,000 American men, babies born lighter than 5.5 pounds had a relative risk of adult hypertension of 1.26 and of type 2 diabetes of 1.75 compared with babies of average birthweight. Similarly, the relative risk of hypertension in light normal babies was 1.43 in 71,000 female U.S. nurses [5, 6].

PROGRAMMING

To explain these findings, the concept of early life physiological “programming” has been proposed [3, 17, 18]. Such programming has been shown in a variety of experimental model systems and reflects the action of a factor during a sensitive period or “window” of development to exert organizational effects that persist through life. Perinatally, programming factors might produce adaptations that optimize survival under conditions of stress or deprivation; such responses might be detrimental when the adult environment is not as adverse as “anticipated.” Of course, genetic and epigenetic factors that restrain fetal growth and produce later disease may also explain these phenomena, and indeed, loci linked to both low birthweight and adult disorders have been

Key words: placenta, fetal growth, glucocorticoid receptor, type 2 diabetes mellitus, hypertension.

© 2000 by the International Society of Nephrology

reported [19, 20]. However, the data suggest that environmental effects occur, and these have been the main focus of mechanistic research.

A major environmental factor advocated in explanation of fetal programming is maternal malnutrition. Indeed, dietary restriction during pregnancy in rats, particularly of protein, produces some reduction in birthweight and permanent hypertension and hyperglycemia in the adult offspring [21–24], effects amplified by later obesity. However, the mechanisms that mediate the effects of maternal undernutrition on later disease in the offspring are not fully understood, and any relevance in modern human populations is moot [25]. Even studies of people born during the extreme starvation of the siege of Leningrad in World War II do not support any strong association between maternal nutrition and offspring disorders [26].

GLUCOCORTICOID PROGRAMMING

Long-term organizational effects are typically found with hormones, particularly steroids. An example is the action of androgens, which neonatally program the expression hepatic steroid-metabolizing enzymes, the development of sexually dimorphic structures in the brain and of sexual behavior [27, 28]. These effects can only be exerted during a specific perinatal period, but then persist throughout life, largely irrespective of any subsequent sex steroid manipulations.

Physiological glucocorticoids (cortisol in humans, corticosterone in rats and mice) are synthesized in the adrenal cortex. Several features of fetal glucocorticoid overexposure suggest a plausible role in the early life programming of adult cardiovascular and metabolic disorders.

Exogenous glucocorticoids retard fetal growth in humans and experimental animals, including nonhuman primates [18, 29–32]. In humans, fetal cortisol levels are increased in intrauterine growth retardation, whether idiopathic or caused by pre-eclampsia [33, 34].

Prenatal glucocorticoids alter the rate of maturation of various organs such as the lung, heart, kidney, and gut [reviewed in 18]. Some effects are transient, whereas others persist throughout life. Perinatal glucocorticoids or stress program specific effects in the brain, notably the hypothalamic-pituitary-adrenal (HPA) axis and dopaminergic motor systems [35–40]. Glucocorticoids also program the immune system [41] and the kidney [42, 43].

Glucocorticoid receptors (GRs) are highly expressed in most, if not all, fetal tissues from midgestation or earlier [44, 45], as well as in placenta and fetal membranes.

Glucocorticoids increase blood pressure and blood glucose levels in adults [46]. Cortisol also elevates fetal blood pressure when infused directly in utero in sheep [47] and at birth in humans and sheep [32, 48].

GLUCOCORTICOID PROGRAMMING

Crucially, prenatal glucocorticoid exposure produces permanently elevated offspring blood pressure levels in later life. Treatment of pregnant rats with modest doses of dexamethasone, a synthetic glucocorticoid used in obstetric practice, reduces birthweight and elevates blood pressure in the adult offspring [49]. Even short-term glucocorticoid exposure in the last trimester increases blood pressure in adult life in rats [50]. These effects are not related to persisting differences in offspring weight, which normalizes by weaning. Last-trimester administration of dexamethasone also “programs” permanent hyperglycemia and, particularly, hyperinsulinemia in the adult offspring [51]. In contrast, earlier prenatal or postnatal exposure to dexamethasone, while reducing fetal or infant growth, has no persisting effects on glucose homeostasis in adult life.

PHYSIOLOGICAL GLUCOCORTICOID AND PLACENTAL 11 β -HYDROXYSTEROID DEHYDROGENASE TYPE 2

Although glucocorticoids are highly lipophilic and rapidly cross the placenta, normally the fetus has much lower levels of physiological glucocorticoids than its mother [52, 53]. This is achieved by placental 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2), which catalyzes the rapid metabolism of cortisol and corticosterone to inert 11-keto forms (cortisone, 11-dehydrocorticosterone) [54, 55]. This placental enzymic barrier ensures that most, but not all [56], maternal cortisol is inactivated so that the majority of cortisol in the human fetal circulation at term is derived from the fetal adrenals. Dexamethasone, a poor substrate for 11 β -HSD2, crosses the placenta readily. The efficiency of placental 11 β -HSD2 near term varies considerably in both rats and humans [49, 57]. It has therefore been hypothesized that relative deficiency of placental 11 β -HSD2, by allowing increased access of maternal glucocorticoids to the fetus, may retard growth and program responses leading to later disease (Fig. 1) [17]. Indeed, in rats, lower placental 11 β -HSD activity and presumably greater fetal exposure to maternal glucocorticoids are seen in the smallest fetuses with the largest placentas. Similar associations between birthweight and placental 11 β -HSD2 have been mooted in humans [57], although not all studies have confirmed this observation [58]. However, deleterious mutations of the 11 β -HSD2 gene in humans associate with very low birthweight [59]. Furthermore, biochemical markers of fetal exposure to glucocorticoids correlate with placental 11 β -HSD2 function at term [60]. Mechanistic studies in pregnant rats with the 11 β -HSD carbenoxolone have shown similar effects to dexamethasone, with reduced birthweight and hypertension and hyperglycemia in the adult offspring (Fig. 2) [61, 62]. These effects of carben-

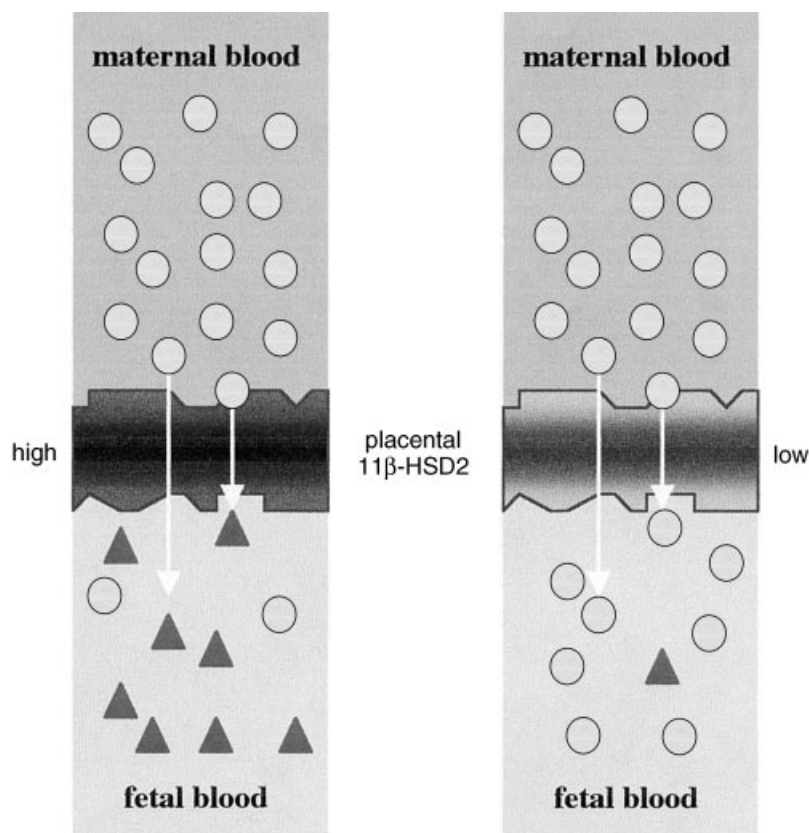


Fig. 1. Placental 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2) acts as a "barrier" to active glucocorticoids (cortisol, ○) in the maternal circulation, converting these to inert 11 keto forms (cortisone, ▲). Lower activity of placental 11 β -HSD2 (right panel) allows greater passage of cortisol to the fetal circulation, which may underlie deleterious short- and long-term effects, including intrauterine growth retardation and "programming" of responses.

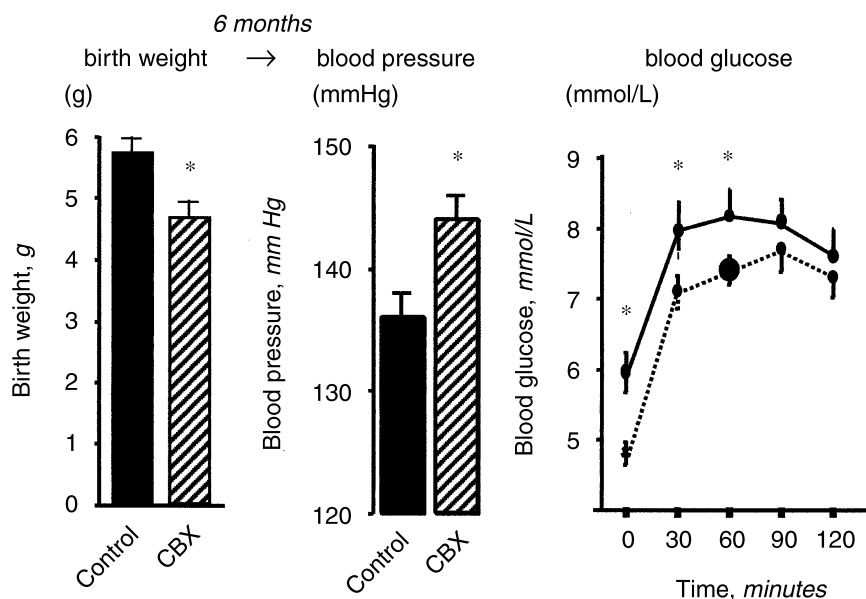


Fig. 2. Prenatal carbenoxolone (CBX) treatment reduces birthweight (▨) and programs permanent hypertension and hyperglycemia (fasting and post-prandial) in the adult offspring compared with control offspring (Con; ■) of vehicle-treated pregnancies. Adult offspring were given oral glucose after 0 minutes. Symbols are: (●, solid line) CBX in utero; (●, dashed line) control.

oxolone are apparently independent of changes in maternal blood pressure or electrolytes, but require maternal glucocorticoids (that is, they are not seen in the offspring of adrenalectomized pregnant rats given carbenoxolone). Intriguingly, dietary protein restriction during rat preg-

nancy selectively attenuates 11 β -HSD2, but not other enzymes in the placenta [63], and leads to hypertension, hyperglycemia, and so forth. Thus, glucocorticoid exposure may be one of a limited series of common mechanisms linking maternal environmental factors with fetal

growth and programming. Indeed, in the maternal protein restriction model, offspring hypertension can be prevented by giving the pregnant dam (and her offspring) glucocorticoid synthesis inhibitors, and can be recreated by concurrent “replacement” of corticosterone, at least in female offspring [64]. A note of caution is necessary, however, since 11 β -HSD2 null mice have a normal birthweight [65]. However, in mice, there is early “shut-off” of 11 β -HSD-2 gene expression in the placenta during midgestation [66], whereas birthweight predominantly reflects growth late in gestation. Thus, in this species, the “barrier” may not exist during the phase of maximal growth. In contrast, the activity of 11 β -HSD2 in rat placenta is maintained until later in gestation [49], and in many other mammals, including humans, placental 11 β -HSD2 activity is fully maintained or even increases toward term [57, 67]. Thus, there are clear species differences in the ontogeny of 11 β -HSD-2 expression in the placenta, hindering the confidence in extrapolating data from results with 11 β -HSD inhibition/disruption in rodent species (particularly the mouse) to humans.

Sites of glucocorticoid action

Glucocorticoids or 11 β -HSD inhibitors administered during pregnancy might affect the fetus, the placenta, and/or the mother. Each is plausible [reviewed in 18], although direct feto-placental effects seem most likely [61, 62]. Nevertheless, maternal hypertension is linked to fetal high blood pressure and hyperglycemia [68, 69]. 11 β -HSD2 is also highly expressed in many fetal tissues until midgestation in rodents and humans [66, 70], which may provide additional tissue-specific controls of steroid action. Finally, several important placental functions are affected by glucocorticoids, including peptide and steroid biosynthesis and antagonism of progesterone action [71–73], which might affect placental growth.

Tissue mechanisms of fetal programming

Prenatal maternal protein restriction or glucocorticoid exposure affects glucose-insulin homeostasis in the adult offspring. A key target appears to be the liver, where glucocorticoids regulate several important processes, including key enzymes regulating carbohydrate and fat metabolism, such as phosphoenolpyruvate carboxykinase, the rate-limiting step in gluconeogenesis. Prenatal glucocorticoid administration programs increased phosphoenolpyruvate carboxykinase gene transcription selectively in the periportal zone of the liver acinus and hence increased enzyme activity [51]. Recent data suggest that permanently increased hepatic GR expression, again only in the periportal zone, may underpin the programming of elevated hepatic phosphoenolpyruvate carboxykinase and adult hyperglycemia. Indeed, rats exposed to dexamethasone in the last trimester show increased rises in plasma glucose levels on exposure to

corticosterone, suggesting the increased hepatic GR expression is of functional importance [51]. Of course, this begs the question of what happens to plasma corticosterone levels in such models.

Prenatal dexamethasone permanently attenuates GR (and mineralocorticoid receptor) gene expression in the adult rat hippocampus, a key region responsible for mediating glucocorticoid negative feedback on the HPA axis. This may well underpin the documented increases in basal plasma corticosterone levels in adulthood by reducing sensitivity to glucocorticoid feedback [50]. Of course, elevated glucocorticoid levels might contribute directly to hypertension and hyperglycemia [49]. The reader will have noted that GR transcript levels are elevated in the liver, but reduced in the hippocampus in the prenatal dexamethasone model. The molecular mechanisms are under current study, but may reflect differential promoter usage by the GR gene in each tissue.

Crucially, the rat models have allowed further predictions to be made in humans. Indeed, low birthweight also correlates with increased adult cortisol levels in humans [74]. Thus, the prenatal glucocorticoid exposure models have reproduced and indeed extended our understanding of the fetal origins phenomena. Whether glucocorticoids and feto-placental 11 β -HSD2 deficiency are common or exceptional mechanisms of fetal programming in humans remains a key question for future investigation.

ACKNOWLEDGMENTS

Research in the authors' laboratory is generously supported by a Wellcome Trust Programme grant (J.R.S.), a Wellcome Veterinary Training Fellowship (M.C.), a WHO Training Fellowship (M.J.N.), and project grants from the Wellcome Trust, the Medical Research Council, the Scottish Hospital Endowments Research Trust, and the High Blood Pressure Foundation.

Reprint requests to Dr. Jonathan R. Seckl, Molecular Medicine Center, University of Edinburgh, Western General Hospital, Edinburgh EH4 2XU, Scotland, United Kingdom.
E-mail: J.Seckl@ed.ac.uk

REFERENCES

1. BARKER DJP, WINTER PD, OSMOND C, MARGETTS B, SIMMONDS SJ: Weight in infancy and death from ischaemic heart disease. *Lancet* 2:577–580, 1989
2. BARKER DJP: Fetal and Infant Origins of Adult Disease, London, British Medical Journal Publishing, 1991, pp 1–343
3. BARKER DJP, GLUCKMAN PD, GODFREY KM, HARDING JE, OWENS JA, ROBINSON JS: Fetal nutrition and cardiovascular disease in adult life. *Lancet* 341:938–941, 1993
4. BARKER DJP: The fetal origins of hypertension. *J Hypertens* 14(Suppl):S117–S120, 1996
5. CURHAN GC, CHERTOW GM, WILLETT WC, SPIEGELMAN D, COLDITZ GA, MANSON JE, SPEIZER FE, STAMPFER MJ: Birth-weight and adult hypertension and obesity in women. *Circulation* 94:1310–1315, 1996
6. CURHAN GC, WILLETT WC, RIMM EB, SPIEGELMAN D, ASCHERIO AL, STAMPFER MJ: Birth weight and adult hypertension, diabetes mellitus, and obesity in US men. *Circulation* 94:3246–3250, 1996
7. FALL CHD, OSMOND C, BARKER DJP, CLARK PMS, HALES CN, STIRLING Y, MEADE TW: Fetal and infant growth and cardiovascular risk factors in women. *Br Med J* 310:428–432, 1995

8. RICHEDWARDS JW, STAMPFER MJ, MANSON JE, ROSNER B, HANKINSON SE, COLDITZ GA, WILLETT WC, HENNEKENS CH: Birth weight and risk of cardiovascular disease in a cohort of women followed up since 1976. *Br Med J* 315:396–400, 1997
9. LEON DA, KOUPILOVA I, LITHELL HO, BERGLUND L, MOHSEN R, VAGERO D, LITHELL UB, McKEIGUE PM: Failure to realise growth potential in utero and adult obesity in relation to blood pressure in 50 year old Swedish men. *Br Med J* 312:401–406, 1996
10. LITHELL HO, McKEIGUE PM, BERGLUND L, MOHSEN R, LITHELL UB, LEON DA: Relation of size at birth to non-insulin dependent diabetes and insulin concentrations in men aged 50–60 years. *Br Med J* 312:406–410, 1996
11. MOORE VM, MILLER AG, BOULTON TJC, COCKINGTON RA, CRAIG IH, MAGAREY AM, ROBINSON JS: Placental weight, birth measurements, and blood pressure at age 8 years. *Arch Dis Child* 74:538–541, 1996
12. FORSEN T, ERIKSSON JG, TUOMILEHTO J, TERAMO K, OSMOND C, BARKER DJP: Mother's weight in pregnancy and coronary heart disease in a cohort of Finnish men: Follow up study. *Br Med J* 315:837–840, 1997
13. YAJNIK CS, FALL CHD, VAIDYA U, PANDIT AN, BAVDEKAR A, BHAT DS, OSMOND C, HALES CN, BARKER DJP: Fetal growth and glucose and insulin metabolism in four-year-old Indian children. *Diabet Med* 12:330–336, 1995
14. LEVINE RS, HENNEKENS CH, JESSE MJ: Blood pressure in prospective population based cohort of newborn and infant twins. *Br Med J* 308:298–302, 1994
15. OSMOND C, BARKER DJP, WINTER PD, FALL CHD, SIMMONDS SJ: Early growth and death from cardiovascular disease in women. *Br Med J* 307:1524–1527, 1993
16. BARKER DJP, BULL AR, OSMOND C, SIMMONDS SJ: Fetal and placental size and risk of hypertension in adult life. *Br Med J* 301:259–263, 1990
17. EDWARDS CRW, BENEDIKTSSON R, LINDSAY R, SECKL JR: Dysfunction of the placental glucocorticoid barrier: A link between the foetal environment and adult hypertension? *Lancet* 341:355–357, 1993
18. SECKL JR: Physiologic programming of the fetus. *Clin Perinatol* 25:939–964, 1998
19. HATTERSLEY A, BEARDS F, BALLANTYNE E, APPLETON M, HARVEY R, ELLARD S: Mutations in the glucokinase gene of the fetus result in reduced birth weight. *Nat Genet* 19:268–270, 1998
20. DUNGER D, ONG K, HUXTABLE S, SHERRIFF A, WOODS K, AHMED M, GOLDING J, PEMBREY M, RING S, BENNETT S, TODD J: Association of the INS VNTR with size at birth. *Nat Genet* 19:98–100, 1998
21. LANGLEY SC, JACKSON AA: Increased systolic blood pressure in adult rats induced by fetal exposure to maternal low protein diets. *Clin Sci* 86:217–222, 1994
22. LANGLEY-EVANS SC, PHILLIPS GJ, JACKSON AA: In utero exposure to maternal low protein diets induces hypertension in weanling rats independently of maternal blood pressure changes. *Clin Nutr* 13:319–324, 1994
23. PETRY CJ, OZANNE SE, WANG CL, HALES CN: Early protein restriction and obesity independently induce hypertension in 1-year-old rats. *Clin Sci* 93:147–152, 1997
24. WOODALL SM, JOHNSTON BM, BREIER BH, GLUCKMAN PD: Chronic maternal undernutrition in the rat leads to delayed postnatal growth and elevated blood pressure of offspring. *Pediatr Res* 40:438–443, 1996
25. MATHEWS F, YUDKIN P, NEIL A: Influence of maternal nutrition on outcome of pregnancy: Prospective cohort study. *Br Med J* 319:339–343, 1999
26. STANNER SA, BULMER K, ANDRES C, LANTSEVA OE, BORODINA V, POTEEN VV, YUDKIN JS: Does malnutrition in utero determine diabetes and coronary heart disease in adulthood? Results from the Leningrad siege study, a cross sectional study. *Br Med J* 315:1342–1348, 1997
27. ARAI Y, GORSKI RA: Critical exposure time for androgenization of the developing hypothalamus in the female rat. *Endocrinology* 82:1010–1014, 1968
28. GUSTAFSSON JA, STENBERG A: Neonatal programming of androgen responsiveness of liver of adult rats. *J Biol Chem* 249:719–723, 1974
29. REINISCH JM, SIMON NG, KARWO WG, GANDELMAN R: Prenatal exposure to prednisone in humans and animals retards intra-uterine growth. *Science* 202:436–438, 1978
30. NOVY MJ, WALSH SW: Dexamethasone and estradiol treatment in pregnant rhesus macaques: Effects on gestation length, maternal plasma hormones and fetal growth. *Am J Obstet Gynecol* 145:920–930, 1983
31. MOSIER HD JR, DEARDEN LC, JANSONS RA, ROBERTS RC, BIGGS CS: Disproportionate growth of organs and body weight following glucocorticoid treatment of the rat fetus. *Dev Pharmacol Ther* 4:89–105, 1982
32. BERRY LM, POLK DH, IKEGAMI M, JOBE AH, PADBURY JF, ERVIN MG: Preterm newborn lamb renal and cardiovascular responses after fetal or maternal antenatal betamethasone. *Am J Physiol* 41:R1972–R1979, 1997
33. GOLAND RS, JOZAK S, WARREN WB, CONWELL IM, STARK RI, TROPPER PJ: Elevated levels of umbilical cord plasma corticotropin-releasing hormone in growth-retarded fetuses. *J Clin Endocrinol Metab* 77:1174–1179, 1993
34. GOLAND RS, TROPPER PJ, WARREN WB, STARK RI, JOZAK SM, CONWELL IM: Concentrations of corticotropin-releasing hormone in the umbilical-cord blood of pregnancies complicated by pre-eclampsia. *Reprod Fertil Dev* 7:1227–1230, 1995
35. MEANEY MJ, AITKEN DH, VIAU V, SHARMA S, SARRIEAU A: Neonatal handling alters adrenocortical negative feedback sensitivity and hippocampal type II glucocorticoid receptor binding in the rat. *Neuroendocrinology* 50:597–604, 1989
36. HENRY C, KABBAJ M, SIMON H, LE MOAL M, MACCARI S: Prenatal stress increases the hypothalamo-pituitary-adrenal axis response in young and adult rats. *J Neuroendocrinol* 6:341–345, 1994
37. LEVINE S: Infantile experience and resistance to physiological stress. *Science* 126:405–406, 1957
38. CATALANI A, MARINELLI M, SCACCIANOCCE S, NICOLAI R, MUSCOLO LAA, PORCU A, KORANYI L, PIAZZA PV, ANGELUCCI L: Progeny of mothers drinking corticosterone during lactation has lower stress-induced corticosterone secretion and better cognitive performance. *Brain Res* 624:209–215, 1993
39. DIAZ R, FUXE K, OGREN SO: Prenatal corticosterone treatment induces long-term changes in spontaneous and apomorphine-mediated motor activity in male and female rats. *Neuroscience* 81:129–140, 1997
40. WEINSTOCK M, MATLINA E, MAOR GI, ROSEN H, McEWEN BS: Prenatal stress selectively alters the reactivity of the hypothalamic-pituitary adrenal system in the female rat. *Brain Res* 595:195–200, 1992
41. REDEI E, HALASZ I, LI LF, PRYSTOWSKY MB, AIRD F: Maternal adrenalectomy alters the immune and endocrine functions of fetal alcohol-exposed male offspring. *Endocrinology* 133:452–460, 1993
42. Deleted in proof
43. CELSI G, KISTNER A, AIZMAN R, EKLOF AC, CECCATELLI S, DESANTIGO A, JACOBSON SH: Prenatal dexamethasone causes oligonephronia, sodium retention, and higher blood pressure in the offspring. *Pediatr Res* 44:317–322, 1998
44. DIAZ R, BROWN RW, SECKL JR: Ontogeny of mRNAs encoding glucocorticoid and mineralocorticoid receptors and 11 β -hydroxysteroid dehydrogenases in prenatal rat brain development reveal complex control of glucocorticoid action. *J Neurosci* 18:2570–2580, 1998
45. COLE TJ, BLENDY JA, MONAGHAN AP, SCHMID W, AGUZZI A, SCHUTZ G: Molecular genetic analysis of glucocorticoid signaling during mouse development. *Steroids* 60:93–96, 1995
46. TONOLO G, FRASER R, CONNELL JMC, KENYON CJ: Chronic low-dose infusions of dexamethasone in rats: Effect on blood pressure, body weight and plasma atrial natriuretic peptide. *J Hypertens* 6:25–31, 1988
47. TANGALAKIS K, LUMBERS ER, MORITZ KM, TOWSTOLESS MK, WINTOUR EM: Effect of cortisol on blood pressure and vascular reactivity in the ovine fetus. *Exp Physiol* 77:709–717, 1992
48. KARI MA, HALLMAN M, ERONEN M, TERAMO K, VIRTANEN M, KOIVISTO M, IKONEN RS: Prenatal dexamethasone treatment in conjunction with rescue therapy of human surfactant: A randomized placebo-controlled multicenter study. *Pediatrics* 93:730–736, 1994

49. BENEDIKTSSON R, LINDSAY R, NOBLE J, SECKL JR, EDWARDS CRW: Glucocorticoid exposure in utero: A new model for adult hypertension. *Lancet* 341:339–341, 1993
50. LEVITT N, LINDSAY RS, HOLMES MC, SECKL JR: Dexamethasone in the last week of pregnancy attenuates hippocampal glucocorticoid receptor gene expression and elevates blood pressure in the adult offspring in the rat. *Neuroendocrinology* 64:412–418, 1996
51. NYIRENDA MJ, LINDSAY RS, KENYON CJ, BURCHELL A, SECKL JR: Glucocorticoid exposure in late gestation permanently programmes rat hepatic phosphoenolpyruvate carboxykinase and glucocorticoid receptor expression and causes glucose intolerance in adult offspring. *J Clin Invest* 101:2174–2181, 1998
52. BEITENS IZ, BAYARD F, ANCES IG, KOWARSKI A, MIGEON CJ: The metabolic clearance rate, blood production, interconversion and transplacental passage of cortisol and cortisone in pregnancy near term. *Pediatr Res* 7:509–519, 1973
53. CAMPBELL AL, MURPHY BEP: The maternal-fetal cortisol gradient during pregnancy and at delivery. *J Clin Endocrinol Metab* 45:435–440, 1977
54. MURPHY BEP, CLARK SJ, DONALD IR, PINSKY M, VEDADY DL: Conversion of maternal cortisol to cortisone during placental transfer to the human fetus. *Am J Obstet Gynecol* 118:538–541, 1974
55. LOPEZ-BERNAL A, FLINT APF, ANDERSON ABM, TURNBULL AC: 11 β -Hydroxysteroid dehydrogenase activity (E.C.1.1.1.146) in human placenta and decidua. *J Steroid Biochem* 13:1081–1087, 1980
56. BENEDIKTSSON R, CALDER AA, EDWARDS CRW, SECKL JR: Placental 11 β -hydroxysteroid dehydrogenase type 2 is the placental barrier to maternal glucocorticoids: Ex vivo studies. *Clin Endocrinol* 46:161–166, 1997
57. STEWART PM, ROGERSON FM, MASON JI: Type 2, 11 β -hydroxysteroid dehydrogenase messenger RNA and activity in human placenta and fetal membranes: its relationship to birth weight and putative role in fetal steroidogenesis. *J Clin Endocrinol Metab* 80:885–890, 1995
58. ROGERSON FM, KAYES KM, WHITE PC: Variation in placental type 2, 11 β -hydroxysteroid dehydrogenase activity is not related to birth weight or placental weight. *Mol Cell Endocrinol* 128:103–109, 1997
59. DAVE-SHARMA S, WILSON RC, HARBISON MD, NEWFIELD R, AZAR M, KROZOWSKI ZS, FUNDER JW, SHACKLETON CHL, BRADLOW HL, WEI JQ, HERTECANT J, MORAN A, NEIBERGER RE, BALFE JW, FATTAH A, DANEMAN D, AKKURT HI, DESANTIS C, NEW MI: Extensive personal experience: Examination of genotype and phenotype relationships in 14 patients with apparent mineralocorticoid excess. *J Clin Endocrinol Metab* 83:2244–2254, 1998
60. BENEDIKTSSON R, BRENNAND J, TIBI L, CALDER AA, SECKL JR, EDWARDS CRW: Fetal osteocalcin levels are related to placental 11 β -hydroxysteroid dehydrogenase activity. *Clin Endocrinol* 42:551–555, 1995
61. LINDSAY RS, LINDSAY RM, EDWARDS CRW, SECKL JR: Inhibition of 11 β -hydroxysteroid dehydrogenase in pregnant rats and the programming of blood pressure in the offspring. *Hypertension* 27:1200–1204, 1996
62. LINDSAY RS, LINDSAY RM, WADDELL B, SECKL JR: Programming of glucose tolerance in the rat: Role of placental 11 β -hydroxysteroid dehydrogenase. *Diabetologia* 39:1299–1305, 1996
63. LANGLEY-EVANS SC, PHILIPS G, BENEDIKTSSON R, GARDNER D, EDWARDS CRW, JACKSON AA, SECKL JR: Maternal dietary protein restriction, placental glucocorticoid metabolism and the programming of hypertension. *Placenta* 17:169–172, 1996
64. LANGLEY-EVANS SC: Hypertension induced by foetal exposure to a maternal low-protein diet, in the rat, is prevented by pharmacological blockade of maternal glucocorticoid synthesis. *J Hypertens* 15:537–544, 1997
65. KOTELEVTSYEV Y, BROWN RW, FLEMING S, KENYON CJ, EDWARDS CRW, SECKL JR, MULLINS JJ: Hypertension in mice lacking 11 β -hydroxysteroid dehydrogenase type 2. *J Clin Invest* 103:683–689, 1999
66. BROWN RW, DIAZ R, ROBSON AC, KOTELEVTSYEV Y, MULLINS JJ, KAUFMAN MH, SECKL JR: The ontogeny of 11 β -hydroxysteroid dehydrogenase type 2 and mineralocorticoid receptor gene expression reveal intricate control of glucocorticoid action in development. *Endocrinology* 137:794–797, 1996
67. PEPE GJ, BABISCHKIN JS, BURCH MG, LEAVITT MG, ALBRECHT ED: Developmental increase in expression of the messenger ribonucleic acid and protein levels of 11 β -hydroxysteroid dehydrogenase types 1 and 2 in the baboon placenta. *Endocrinology* 137:5678–5684, 1996
68. HIMMELMANN A, SVENSSON A, HANSSON L: 5 Year follow up of blood pressure and left ventricular mass in children with different maternal histories of hypertension: The hypertension in pregnancy offspring study. *J Hypertens* 12:89–95, 1994
69. HIMMELMANN A, HIMMELMANN K, SVENSSON A, HANSSON L: Glucose and insulin levels in young subjects with different maternal histories of hypertension: The hypertension in pregnancy offspring study. *J Intern Med* 241:19–22, 1997
70. STEWART PM, MURRY BA, MASON JI: Type 2, 11 β -hydroxysteroid dehydrogenase in human fetal tissues. *J Clin Endocrinol Metab* 78:1529–1532, 1994
71. RINGLER GE, KALLEN CB, STRAUSS JFI: Regulation of human trophoblast function by glucocorticoids: Dexamethasone promotes increased secretion of chorionic gonadotropin. *Endocrinology* 124:1625–1631, 1989
72. STEELE PA, FLINT APF, TURNBULL AC: Activity of steroid C-17,20-lyase in the ovine placenta: Effect of exposure to foetal glucocorticoid. *J Endocrinol* 69:239–246, 1976
73. KARALIS K, GOODWIN G, MAJZOUB JA: Cortisol blockade of progesterone: A possible molecular mechanism involved in the initiation of human labor. *Nat Med* 2:556–560, 1996
74. PHILLIPS DI, FALL CHD, WHORWOOD CB, SECKL JR, WOOD PJ, BARKER DJP, WALKER BR: Elevated plasma cortisol concentrations: An explanation for the relationship between low birth weight and adult cardiovascular risk factors. *J Clin Endocrinol Metab* 83:757–760, 1998